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(54) Title: GENETIC SEQUENCES, DIAGNOSTIC AND/OR QUANTIFICATION METHODS AND DEVICES FOR THE IDENTIFICATION OF *STAPHYLOCOCCI* STRAINS

(57) Abstract

The present invention is related to oligonucleotides for the specific identification of *Staphylococci* species which nucleotide sequence has between 15 and 350 base pairs, preferably between 15 and 45 base pairs, obtained from the "consensus" *femA* nucleotide sequence (CNS) of the figure or its complementary strand. The present invention is also related to a method and a diagnostic device using said oligonucleotide for the identification of various types of *Staphylococci* species strains.

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GENETIC SEQUENCES, DIAGNOSTIC AND/OR QUANTIFICATION METHODS
10 AND DEVICES FOR THE IDENTIFICATION OF STAPHYLOCOCCI STRAINS

Field of the invention

15 The present invention refers to new genetic sequences, diagnostic and/or quantification methods and devices using said sequences for the identification of various types of *Staphylococci* strains as well as the therapeutical aspects of said genetic sequences.

Background of the invention

20 Increasing incidence of nosocomial infections by multiresistant bacteria (even to antibiotics like vancomycin) is a world-wide concern. Methicillin-resistant coagulase-negative *Staphylococci* (MR-CNS) and *S. aureus* (MRSA) express a high level cross-resistance to all β -lactam antibiotics (Ryffel et al. (1990), Refsahl et al. 25 (1992)). They have an additional low-affinity penicillin-building protein, PBP2a (PBP2'), encoded by the *mecA* gene. The *mecA* determinant is found in all multiresistant staphylococcal species (Chackbart et al. (1989), Suzuki et 30 al. (1992), Vannuffel et al. (1995)) and is highly conserved among the different species (Ryffel et al. (1990)).

Several other chromosomal sites, in which transposon inactivation reduces the level of β -lactam resistance, have been identified in *S. aureus* (SA) (Hiramatsu (1992), Berger-Bächi et al. (1992), de Lancastre 5 et al. (1994)). The appropriate functioning of these regulator genes rather than the quantity of PBP2a determines the minimal inhibitory concentration value and homogeneous expression of resistance of staphylococcal isolates (Ryffel et al. (1994), de Lancastre et al. 10 (1994)).

The *femA-femB* operon, initially identified in *S. aureus*, is one of those genetic factors essential for methicillin resistance (Berger-Bächi et al. (1989)). It is involved in the formation of the characteristic 15 pentaglycine side chain of the SA peptidoglycan (Stranden et al. (1997)). Unlike other regulatory genes, *femA* was shown to retain a strong conservation over time in clinical isolates of MRSA, hence confirming its key role in cell wall metabolism and methicillin resistance (Hurlimann-Dalel 20 et al. (1992)). In contrast to *mecA*, *femA-femB* is present both in the genome of resistant and susceptible SA strains (Unal et al. (1992), Vannuffel et al. (1995)).

Often, identification of the *Staphylococci* is limited to a rapid screening test for *S. aureus*, and non-*S. aureus* isolates are simply reported as coagulase-negative 25 *Staphylococci*. In fact, these bacteria isolates include a variety of species and many different strains (Kleeman et al. (1993)). There is little epidemiological information related to the acquisition and spread of these organisms. 30 This is potentially due to the lack of an easy and accurate way to identify species and to provide clinically timely informations.

Several molecular assays designed for detecting *femA* in SA failed to amplify an homologous sequence in coagulase-negative *Staphylococci* (Kizaki et al. (1994), Vannuffel et al. (1995)). Nevertheless, low-5 stringency heterologous hybridisation analysis suggested the presence of such a structurally related gene in *S. epidermidis* (SE) (Unal et al. (1992)).

These data were followed by complete identification and sequence analysis of the *femA* and *femB* 10 open reading frames in *S. epidermidis* (Alborn et al. (1996)). Intra- and interspecies relatedness of these genes and conservation of genomic organisation are therefore consistent with gene duplication of one of these genes in an ancestral organism and the possibility of *femA* 15 phylogenetic conservation in all staphylococcal species (Alborn et al. (1996)).

The complete genetic sequence of the *femA* gene of *S. epidermidis*, the protein encoded by the *femA* gene (*FemA*) and vectors and micro-organisms comprising 20 genes encoding the *FemA* protein are described in the US patent 5,587,307.

Aims of the invention

The present invention aims to provide new 25 genetic sequences, methods and devices for the improvement of the identification and/or the quantification of various types of *Staphylococci* strains through their *femA*-like determinants, which allow by a rapid screening their epidemiological study.

30 Another aim of the invention is to identify similar genetic sequences which may exist in known or not

known *Staphylococci* species or other gram-positive bacterial strains.

A last aim of the present invention is to provide new sequences encoding *femA* proteins of 5 *Staphylococci* species, their *femA* proteins, vector(s) comprising said nucleotide sequences and cell (s) transformed by said vector(s) for possible therapeutical applications.

10 Summary of the invention

The Inventors have identified new DNA and amino acid sequences from new strains of *Staphylococcus hominis*, *Staphylococcus saprophyticus* and *Staphylococcus haemolyticus*. Said new nucleotide sequences allow an 15 alignment of these new sequences with the *femA* gene from *Staphylococci* previously described (*S. aureus*, *S. epidermidis* and *S. saprophyticus*). By the alignment of more than 2 sequences, preferably more than 4 sequences, the Inventors have identified for the first time a consensus 20 *femA* sequence useful for molecular genotyping of different *Staphylococci* species which was not possible previously, when only few *femA* sequences of *Staphylococci* strains were known.

Therefore, a first aspect of the present 25 invention is related to the "consensus" nucleotide sequence as represented in the enclosed Figure 3. With said "consensus" nucleotide sequence, the Inventors were able to provide oligonucleotides (such as primers or probes) which can be used for the genetic amplification, the 30 identification and/or quantification of various *femA* sequences which are specific of known or unknown *Staphylococci* species.

The *femA* sequence is known to be involved with the biosynthesis of glycine-containing cross-bridges of the peptidoglycan and the peptidoglycan organisation is also known to be well conserved among various *Staphylococci* 5 species and possibly among other gram-positive bacteria.

Therefore, it is also possible to use the new "consensus" *femA* sequence and said new oligonucleotides extrapolated from the alignment of the sequences presented in Figure 3, for the molecular genotyping of other 10 *Staphylococci* species and possibly other gram-positive bacteria. It is also known that the *femA* sequence is similar to the *femB* sequence. Therefore, these 15 oligonucleotides could also be used for the molecular genotyping of *femB* genes of different *Staphylococci* species or other gram-positive bacteria.

Another aspect of the present invention concerns the possible therapeutical uses of new *femA* nucleotide sequences isolated from the strains *S. hominis*, *S. saprophyticus*, *S. haemolyticus*, *S. lugdunensis*, *S. 20 xylosus*, *S. capitis*, *S. schleiferi* and *S. sciuri* having a nucleotide or amino acid sequence which presents more than 85%, preferably more than 90% homology or 100% homology with the genetic sequences presented in the Figures 6 to 13, their complementary strand and functional variants 25 thereof. Functional variants of said amino acid sequences are peptides which contain one or more modifications to the primary amino acids sequence and retain the activity of the complete and wild type *femA* molecule. Variants of the peptide are obtained by nucleotidic sequences which differ 30 from the above-identified described sequences by a degeneration of their genetic code or are sequences which hybridise with said sequences or their complementary

strand, preferably under stringent conditions such as the ones described in the document Sambrook et al., §§ 9.47-9.51 in *Molecular Cloning : A Laboratory Manual*, Cold Spring Harbor, Laboratory Press, Cold Spring Harbor, New York (1989).

A further aspect of the present invention concerns the recombinant vector (i.e. constructions into which the sequence of the invention may be inserted for transport in different genetic environments and for expression in a host cell, such as a phagemide, a virus, a plasmid, a cationic vesicle, a liposome, etc.) comprising said nucleotide sequences and their complementary strands, or the corresponding RNA sequences, possibly linked to one or more regulatory sequences or markers (resistance to antibiotics, enzyme coding sequences, ...) active into a cell.

Similarly, the nucleic acid sequence according to the invention may be obtained by synthetic methodology well known by the person skilled in the art, such as the one described by Brown et al. ("Method of Enzymology", Acad. Press, New-York, No. 68 pp. 109-151 (1979)) or by conventional DNA synthesising apparatus such as the applied biosystem model 380A or 380B DNA synthesiser.

Other aspects of the present invention concern the recombinant host (prokaryotic) cell transformed by said vector and the purified (possibly recombinant) proteins or peptides encoded by said nucleic acid sequences, possibly linked to a carrier molecule such as BSA and obtained by said cells. Said recombinant proteins or peptides could be obtained by genetic engineering or could be obtained by synthesis (see US patent 5,587,307

incorporated herein by reference) and may comprise residues enhancing their stability (resistance to hydrolysis by proteases, etc.) such as the one described by Nachman et al. (*Regul. Pept.* Vol. 57, pp. 359-370 (1995)).

5 A preferred vector for expression in a *E. coli* host cell is derived from the *E. coli* plasmid pET-11A available from Novagen Inc. (Catalogue No. 69436-A). The transformation technique used with the above-identified vector has been described in the US Patent 5587307.

10 A further aspect of the present invention concerns the inhibitor (used to possibly treat (with addition of antibiotics) antibiotics resistance bacteria) directed against said proteins, peptides or nucleic acid molecules. Advantageously, said inhibitor is a antibody, 15 preferably a monoclonal antibody, or an antisense nucleotide molecule, such as a ribozyme, which could be present in a vector in order to block the expression of said *femA* nucleotide sequences.

A last aspect of the present invention 20 concerns the pharmaceutical composition, preferably a vaccine, against *Staphylococci* infections in an animal, including a human, comprising a pharmaceutically acceptable carrier and a sufficient amount of an active compound selected from the group consisting of said nucleic acid 25 molecules, vectors, recombinant host cells transformed by said vector(s), inhibitors (directed against said proteins, peptides or nucleic acid molecules) and a mixture thereof.

Another aspect of the present invention concerns oligonucleotides which are (DNA) sequences having 30 between 15 and 350 base pairs, preferably between 17 and 250 base pairs (such as primers or probes) obtained from the consensus sequence of Figure 3 or its complementary

strand. Preferably, said oligonucleotides are primers having between 15 and 45 base pairs, more preferably between 17 and 25 base pairs.

According to a first embodiment of the 5 present invention, said oligonucleotide is a primer having between 15 and 45 base pairs, which presents more than 60%, advantageously more than 70%, preferably more than 80%, more specifically more than 90% homology with (fragments of) the "consensus" *femA* nucleotide sequence (CNS) 10 identified in the Figure 3. ,

Therefore, the oligonucleotides according to the invention are new sequences or preferred fragments of known sequences of *S. aureus*, *S. epidermidis* or *S. simulans* but not the complete wild type known *femA* nucleotide 15 sequence.

Preferably, the oligonucleotide according to the invention is selected from the group consisting of the following nucleotide sequences :

- ANAATGAANTTACNAATTNACNGCNANAGANTT
- 20 and more particularly *femS1* TAATGAAGTTACAAAATTT or *femS2* TAATGAAGTTACNAAAATTT
- ATGNCNNANAGNCATTNACNCANA and more particularly *femU1* ("universal" sequence sense of the multiplex PCR) : TGCCATATAGTCATTTACGC
- 25 - TAGTNGGNATNAANAANAANNATAANGANGTNATTGC
- GTNCCNGTNATGAAANTNTTNAANTANTTTATTTC
- AATGCNGGNANANGATTGG
- GNAANNGNAANACNAAAAAGTNNANAANAATGGNGTNAAAGT and more particularly *fsq1S* (et 1AS) : 30 AAAAAGTTCAAAAAATGG and *fsq2S* (and 2AS) : AAAAAGTACAAAAAATGG
- AAGANGANNTNCCNATNTTNNNGNTCATTNATGGANGATAC

- TATATNNANTTTGATGANTA
- AANGANATNGANAAAANGNCCNGANAANAAAAA
and more particularly *fsq3S* (and 3AS) :
AAAGATATTGAAAAACGA, *fsq4S* (and 4AS) :
AAAGATATTGAAAAGAGACC, *fsq5S* (and 5AS) :
AAAGATATCGAGAAAGAC and *fsq6S* (and 6AS) :
AAAGACATCGACAAGCGT.
- ANCAGGNAANGAATTACCNAT
and more particularly *fem1* (primer for the production
of a probe and of marked amplicons for reverse
hybridisation experiment) : GAACATGGTAATGAATTAC
- AATCCNTNTGAAGTNGTNTANTANGCNGGTGG
- AGNTATGCNNTNCAATGGNNNATGATTAANTATGC
- TTTANNGANGANGCNGAAGATGNNGNGTNNTNAANTTNAAAAA
15 and more particularly *fem3bio* (primer for the
production of a probe and of marked amplicons for
reverse hybridisation experiment) :
TTTACTGAAGATGCTGAAGA
- GTTGGNGANTTNNTNAAACC
20 and more particularly *fem2* (primer for the production
of a probe and of marked amplicons for reverse
hybridisation experiment) : GTTGGTGACTTTATTAAACC
- ATGAAATTTACAGAGTTAA (= *femAS1*)

25 Said primer(s) will be designated hereafter
as "universal primer(s)".

A further aspect of the present invention
concerns the oligonucleotide being either a primer or a
probe as above-described, having between 15 and 350 base
30 pairs, preferably between 17 and 250 base pairs, or a
primer having between 15 and 45 base pairs, more preferably
between 17 and 25 base pairs, which will be designated

hereafter as "specific primer(s)", having a nucleotide sequence which presents less than 50%, advantageously less than 40%, preferably less than 30%, more specifically less than 20% homology with (fragments of) the "consensus" *femA* 5 nucleotide sequence (CNS) identified in the Figure 3 and with another *femA* nucleotide sequence specific for other *Staphylococci* strains.

Advantageously, said "specific primer" is selected from the group consisting of the following 10 nucleotide sequences :

- ACAGCAGATGACATCATT
- TAATGAAAGAAATGTGCTTA
- ACACAACTTCAATTAGAAC
- AGTATTAGCAAATGCGG
- 15 - ATGCATATTTCCGTAA
- CAGCAGATGACATCATT
- CATCTAAAGATATATTAAATGGA
- AGTATTAGCAAATGCGGGTCAC
- CAACACAACTTCAATTAGAA

20

The oligonucleotides according to the invention are selected according to their physiochemical properties in order to avoid cross-hybridisation between themselves. Said primers are not complementary to each 25 other and they contain a similar percentage of bases GC.

Said oligonucleotides are used in an identification and/or quantification method of one or more *Staphylococcus* species and possibly other gram-positive bacteria.

30 Therefore, another aspect of the present invention is related to an identification and/or

quantification method of a *Staphylococci* species which may present resistance to one or more antibiotic(s), and is possibly combined with a method for the identification of a resistance to antibiotics, especially β -lactam antibiotics,
5 (for instance through the identification of a variant of the *mecA* gene as described by Vannuffel et al. (1998)).

The method for the detection, the identification and/or the quantification of a bacteria, preferably a staphylococcal species, comprises the steps
10 of :

- obtaining a nucleotide sequence from said bacteria present in a sample, preferably a biological body sample obtained from a patient such as blood, serum, dialyse liquid or cerebrospinal liquid, or from any other
15 bacteriological growth medium,
- possibly purifying said nucleotide sequence from possible contaminants,
- possibly amplifying by known genetic amplification techniques said nucleotide sequence with one or more
20 universal oligonucleotide(s) (universal primer(s)) according to the invention, and
- identifying the specific gram-positive bacteria species, preferably the specific *Staphylococci* species :
 - by a comparative measure of the length of the
25 (possibly amplified) nucleotide sequence or
 - by reverse hybridisation of the (possibly amplified) nucleotide sequence with one or more specific oligonucleotide(s) (specific probe(s) or primer(s)) according to the invention which are
30 specific of said bacteria, said oligonucleotide(s) being preferably immobilised on a solid support.

The comparative measure of the length of a possibly amplified nucleotide sequences can be performed by the analysis of their migration (compared with a known ladder) upon an electrophoresis gel.

5 Preferably, the genetic amplification technique is selected from the group consisting of PCR (US patent 4,965,188), LCR (Landgren et al., *Sciences*, 241, pp. 1077-1080 (1988)), NASBA (Kievits et al., *J. Virol. Methods*, 35, pp. 273-286 (1991)), CPR (patent WO95/14106) 10 or ICR.

The specific detection of the possibly amplified nucleotide sequences can be obtained by the person skilled in the art by using known specific gel electrophoresis techniques, *in situ* hybridisation, 15 hybridisation on solid support, *in solution*, on dot blot, by Northern blot or Southern blot hybridisation, etc.

Advantageously, the probes which are specific of the bacteria are immobilised on a solid support according to the method described in the international 20 patent application WO98/11253 incorporated herein by reference.

Said specific oligonucleotides (probes or "elongated" primers) have a length comprised between 50 and 350 base pairs, preferably between 120 and 250 base pairs, 25 and are fixed to the solid support by a terminal 5' phosphate upon an amine function of the solid support by carbodiimide reaction (as described in the document WO98/11253 incorporated herein by reference).

The solid support can be selected from the 30 group consisting of cellulose or nylon filters, plastic supports such as 96-well microtiter plates, microbeads,

preferably magnetic microbeads, or any other support suitable for the fixation of a nucleotide sequence.

The method according to the invention can be advantageously combined with another specific detection 5 step of a possible resistance to antibiotics, especially β -lactam antibiotics (for instance through the identification by the above-described technique of variants of the *mecA* gene as described by Vannuffel et al. (1998)).

The present invention concerns also a 10 diagnostic and/or quantification device or kit for the identification and/or the quantification of a *Staphylococcus* species or other gram-positive bacteria, comprising the oligonucleotides according to the invention and possibly all the media necessary for the identification 15 of a (possibly amplified) nucleotide sequence of said bacteria through any one of the above-described methods.

Advantageously, the method and device according to the invention are adapted for the quantification of said *Staphylococci* strains by the use of 20 a "internal or external standard sequence", preferably the one described in the patent application WO98/11253 incorporated herein by reference.

Therefore, according to a first embodiment of the present invention, the nucleic acid sequence from a 25 *Staphylococcus* species, for instance *Staphylococcus aureus*, is amplified by a "universal primer" and by a "specific primer" which is specific for *S. aureus*. The identification of *S. aureus* will be obtained upon an agarose electrophoresis gel wherein the amplified nucleotide 30 sequence (shorter than the amplified nucleotide sequence of another *Staphylococci* species such as *S. epidermidis*) and identified by the use of a comparative ladder.

According to another embodiment of the present invention, a *Staphylococcus* species (such as *S. aureus*) is identified by reverse hybridisation of the amplified nucleotide sequence with a probe which is 5 specific of said bacteria and which is immobilised on a solid support such as filter.

The present invention will be described in details in the following non-limiting examples, in reference to the Figures described hereafter.

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Short description of the drawings

The Figure 1 represents 5 partially overlapping fragments of the *femA* genes from *S. hominis*, *S. saprophyticus* and *S. haemolyticus* obtained 15 by PCR amplification.

The Figure 2 represents the alignment of the nucleotide sequences of *femA* genes from *S. hominis*, *S. saprophyticus*, *S. aureus*, *S. epidermidis* and *S. haemolyticus*.

20 The Figure 3 represents the consensus sequence according to the invention.

The Figure 4 represents the result of differential diagnosis between different strains of *Staphylococci* by reverse hybridisation.

25 The Figure 5 represents amplification of CNS species under universal conditions.

Figures 6 to 13 represent the complete *femA* wild type genetic sequence of the strains *S. hominis*, *S. saprophyticus*, *S. haemolyticus*, *S. lugdunensis*, *S. xylosus*, *S. capitis*, *S. schleiferi* and *S. sciuri*.
30

ExamplesExample 1 : Sequencing strategy

Fragments of the *femA* genes from *S. hominis* and *S. saprophyticus* have been obtained by PCR amplification, in low stringency annealing conditions. Primers used for amplification are matching the potentially conserved regions and have been designed according to sequences homologies between *S. aureus*, *S. saprophyticus* and *S. epidermidis* *femA* nucleotide sequences. For both *S. hominis* and *S. saprophyticus* species, 5 partially overlapping fragments have been synthesised allowing the sequencing of the entire *femA* genes (Fig. 1).

Example 2 : Identification of a consensus sequence

Alignment of the nucleotide sequences of *femA* genes from *S. hominis* and *S. saprophyticus* as well as with *femA* genes sequenced to date from *S. aureus* (GenBank accession number M23918), *S. epidermidis* (GenBank accession number U23713) and *S. haemolyticus* is presented in Fig. 3 and has allowed to propose a "consensus" *femA* nucleotide sequence (CNS) whose genomic organisation displays highly conserved regions flanked by variable ones. On this basis, interspecies phylogenetic variations could be exploited to design genotyping strategies for species-specific identification of *Staphylococci*. The "consensus" sequence is therefore a powerful molecular tool for specific diagnostic of staphylococcal infections.

Example 3 : Sequencing of other staphylococcal *femA* genes

The consensus sequence can be exploited for designing universal primers allowing the production, under permissive annealing conditions, of overlapping PCR

products whose sequencing will identify the entire *femA* sequence.

Example 4 : Differential diagnosis between *S. aureus*, *S. epidermidis*, *S. hominis* and *S. saprophyticus* by reverse hybridisation

The Inventors have set up a reverse hybridisation assay for rapid and combined identification of the most clinically relevant *Staphylococci* species, and 10 their *mecA* status. Two sets of primers, chosen in a conserved domain of the consensus sequence (*bioU1-bioU3* and *fem1-fem3bio*), amplifying a 286 and bio-220 bp fragments, respectively) were synthesised. Species-specificity of *femA* amplicons was insured by the genomic variability between 15 the conserved regions. *FemA* probes were immobilised on nylon strips. Hybridisation was performed with biotinylated *femA* PCR fragments from the strain of interest. The strategy was first assessed with ATCC strains (*S. aureus*, *S. epidermidis*, *S. hominis* and *S. saprophyticus*) (Fig. 4). 20 Specificity was identified by standard methods. Accuracy was 100% for species identification.

Example 5 : Differential diagnosis between staphylococcal species

25 This assay is able to identify any staphylococcal species if following requirements are fulfilled :

- primers *fem1*, *fem2* and *fem3bio* are universal for *Staphylococci*;
- 30 - there is a wide enough phylogenetic variation between any CNS species to promote a specific hybridisation.

The first requirement is fulfilled for, i.e., *S. haemolyticus*, *S. capitis*, *S. cohnii*, *S. xylosus*, *S. simulans*, *S. lugdunensis*, *S. schleiferi* and *S. warneri* strains (Fig. 5).

5

Example 6 : Multiplex amplification of *femA* and *mecA* genetic determinants for a molecular diagnosis of a specific staphylococcal infection

A total of 48 patients treated in 4 10 contiguous intensive care units were included in the study. Endotracheal aspirates (ETA) were collected from the patients and submitted to the multiplex PCR analysis according to the technique described by Vannuffel et al. (1995). Clinical specimens were homogenised in 5 ml of TE 15 buffer (20 mM TRIS HCl, pH 8.0, 10 mM EDTA) containing 2% (w/v) SDS.

The homogenate (1.5 ml) was then centrifuged for 5 minutes at 7500 xg. The cellular pellet was washed once with TE buffer lysed in the presence of 1% (v/v) 20 Triton X-100 and 50 µg of lysostaphin (Sigma) and incubated for 15 minutes at 37 °C. Lysis was completed by adding 100 µg of proteinase K (Boehringer). The lysate was incubated for another 5 minutes at 55 °C and 5 minutes at 95 °C, and centrifuged at 4000 xg for 5 minutes.

25 In order to purify bacterial DNA, 200 µl of supernatant were then filtered on a Macherey-Nagel Nucleospin C+T® column and eluted with 200 µl sterile H₂O. Two different amounts of DNA suspension (2 µl and 200 µl) were submitted to multiplex PCR amplification with the 30 primers 5'-TGGCTATCGTGTACAAATCG-3' and 5'-

CTGGAACTTGTTGAGCAGAG-3' for *mecA* and the above-described primers for *femA*, yielding different fragments.

femA and *mecA* signals were found in specimens containing either susceptible *S. aureus* (n = 10) and 5 methycillin-resistant coagulase-negative *Staphylococci* (n = 6) respectively. On the other hand, no signal was obtained from ETA gram-negative bacteria (n = 5) as well as MS-CNS (n = 6) and from 5 ETA containing normal pharyngeal flora.

10 This multiplex, PCR strategy for detecting *Staphylococci* in ETA was completed in less than 6 hours either on the day of the samples' collection. This is an advantage with respect to the time required to conventional identification and susceptibility tests (48 to 72 hours).

15

Example 7 : Amplification, cloning and sequencing of other *femA* genes

Two primers were selected among the conserved parts of the consensus sequence for the amplification of 20 the *femA* gene.

These primers are *femS1*, *femS2* and *femAS1* (anti-sense primer). ADN from strains of *Staphylococcus hominis*, *saprophyticus*, *haemolyticus*, *lugdunensis*, *schleiferi*, *sciuri*, *xylosus*, *simulans*, *capitis*, *gallinarum*, 25 *cohnii* and *warneri* were amplified from said primers and amplification fragments were cloned in the vector pCR®-XLTOPO and introduced by electroporation in *E. coli* cells TOP10 (TOPO XL PCR Cloning Kit®, Invitrogen, Carlsbad, CA).

Amplified fragments of strain *S. lugdunensis*, 30 *schleiferi*, *sciuri*, *xylosus*, and *capitis* were sequenced by Taq Dye Deoxy Terminator Cycle® sequencing on a ABI 277 DNA

sequencer® (PE Applied Biosystems, Foster City, CA) by the following primers :

femS1 or *femS2* or *femAS1*

fsq1S and *fsq1AS*

5 *fsq2S* and *fsq2AS*

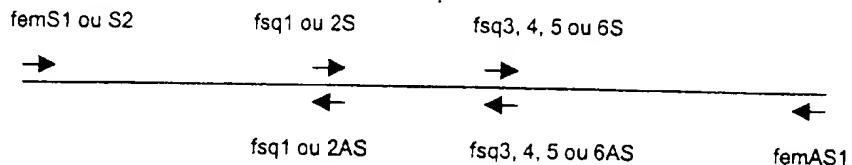
fsq3S and *fsq3AS*

fsq4S and *fsq4AS*

fsq5S and *fsq5AS*

fsq6S and *fsq6AS*

10



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CLAIMS

1. Oligonucleotide for the specific identification of *Staphylococci* species which nucleotide sequence has between 15 and 45 base pairs, preferably 5 between 15 and 25 base pairs, and which presents more than 60% homology with the "consensus" *femA* nucleotide sequence (CNS) of Fig. 3.

2. Oligonucleotide according to claim 1 for the specific identification of *Staphylococci* species, which 10 nucleotide sequence has between 15 and 45 base pairs, preferably between 17 and 25 base pairs, and which presents more than 70% homology with the "consensus" *femA* nucleotide sequence (CNS) of Fig. 3.

3. Oligonucleotide according to claim 1 or 2 15 for the specific identification of *Staphylococci* species, which nucleotide sequence has between 15 and 45 base pairs, preferably between 17 and 25 base pairs, and which presents more than 80% homology with the "consensus" *femA* nucleotide sequence (CNS) of Fig. 3.

20 4. Oligonucleotide according to any of the claims 1 to 3 for the specific identification of *Staphylococci* species, which nucleotide sequence has between 15 and 45 base pairs, preferably between 17 and 25 base pairs, and which presents more than 90% homology with 25 the "consensus" *femA* nucleotide sequence (CNS) of Fig. 3.

5. Oligonucleotide according to any of the preceding claims, which is selected from the group consisting of the following nucleotide sequences :

- ANAATGAANTTTACNAATTNACNGCNANAGANTT
30 and more particularly TAATGAAGTTACAAAATT or
TAATGAAGTTACNAAATT

- ATGNCNNANAGNCATTNACNCANA
and more particularly TGCCATATAGTCATTTACGC
- TAGTNGGNATNAANAANAANNATAANGANGTNATTGC
- GTNCCNGTNATGAAANTNTNAANTANTTTATT
- 5 - AATGCNGGNANGATTGG
- GNAANGNAANACNAAAAAGTNNANAANAATGGNGTNAAAGT
and more particularly AAAAAGTTCAAAAAATGG and
AAAAAGTACAAAAAATGG
- AAGANGANNTNCCNATNTNNNGNTCATTNATGGANGATAC
- 10 - TATATNNANTTGATGANTA
- AANGANATNGANAAANGNCCNGANAANAAAA
and more particularly AAAGATATTGAAAAACGA,
AAAGATATTGAAAAGAGACC, AAAGATATCGAGAAAGAC and
AAAGACATCGACAAGCGT.
- 15 - ANCAGGNAANGAATTACCNAT
and more particularly GAACATGGTAATGAATTAC
- AATCCNTNTGAAGTNGTNTANTANGCNGGTGG
- AGNTATGCNNTNCAATGGNNNATGATTAANTATGC
- TTTANNGANGANGCNGAAGATGNNGNGTNNTNAANTTNAAAAA
- 20 and more particularly TTTACTGAAGATGCTGAAGA
- GTTGGNGANTNNNTNAAACC
and more particularly GTTGGTGACTTTATTAAACC
- ATGAAATTACAGAGTTAA

6. Oligonucleotide for the specific
25 identification of *Staphylococci* species which nucleotide
sequence has between 15 and 350 base pairs, preferably
between 17 and 250 base pairs, and which presents less than
50% homology with the "consensus" *femA* nucleotide sequence
(CNS) of Fig. 3.

30 7. Oligonucleotide according to claim 6 for
the specific identification of *Staphylococci* species which
nucleotide sequence has between 15 and 350 base pairs,

preferably between 17 and 250 base pairs, and which presents less than 40% homology with the "consensus" *femA* nucleotide sequence (CNS) of Fig. 3.

8. Oligonucleotide according to claim 6 or 7
5 for the specific identification of *Staphylococci* species which nucleotide sequence has between 15 and 350 base pairs, preferably between 17 and 250 base pairs, and which presents less than 30% homology with the "consensus" *femA* nucleotide sequence (CNS) of Fig. 3.

10 9. Oligonucleotide according to any of the claims 6 to 8 for the specific identification of *Staphylococci* species which nucleotide sequence has between 15 and 350 base pairs, preferably between 17 and 250 base pairs, and which presents less than 20% homology with the
15 "consensus" *femA* nucleotide sequence (CNS) of Fig. 3.

10. Oligonucleotide according to claim 6, being a primer which nucleotide sequence has between 15 and 45 base pairs, preferably between 17 and 25 base pairs.

11. Oligonucleotide according to claim 10,
20 which is selected from the group consisting of the following nucleotide sequences :

- ACAGCAGATGACATCATT
- TAATGAAAGAAATGTGCTTA
- ACACAACTTCAATTAGAAC
- 25 - AGTATTAGCAAATGCGG
- ATGCATATTTCCGTAA
- CAGCAGATGACATCATT
- CATCTAAAGATATATTAAATGGA
- AGTATTAGCAAATGCGGGTCAC
- 30 - CAACACAACTTCAATTAGAA

12. Identification and/or quantification method of a *Staphylococci* species, which may present resistance to antibiotics and which is present in a sample, said method comprising the steps of :

5 - obtaining a nucleotide sequence from a *Staphylococci* species present in the sample,

- amplifying said nucleotide sequence with one or more oligonucleotide(s) according to the claims 1 to 8, and

- identifying and possibly quantifying the specific

10 *Staphylococci* species :

- by reverse hybridisation of the amplified nucleotide sequence with one or more oligonucleotide(s) according to the claims 9 to 11 which is (are) specific of said *Staphylococci* species and is (are) immobilised on a solid support or
- by a comparative measure of the length of the amplified nucleotide sequence.

13. Diagnostic device for the identification

20 of *Staphylococci* species comprising the oligonucleotide according to any of the preceding claims 1 to 11 and possibly all the media necessary for the identification of an amplified sequence of said *Staphylococci* species through any one of the methods selected from the group consisting

25 of in situ hybridisation, hybridisation on a solid support, in solution on dot blot, Northern blot, Southern blot, probe hybridisation by the use of an isotopic or non-isotopic label, genetic amplification or a mixture thereof.

14. *femA* genetic sequence which presents more

30 than 90% homology with a nucleotide or amino acid sequence selected from the group consisting of the nucleotide or

amino acid sequences represented in the enclosed Fig. 6 to 13.

15. Genetic sequence according to claim 14,
being the nucleotide sequence of Fig. 6.

5 16. Genetic sequence according to claim 14,
being the amino acid sequence of Fig. 6.

17. Genetic sequence according to claim 14,
being the nucleotide sequence of Fig. 7.

18. Genetic sequence according to claim 14,
10 being the amino acid sequence of Fig. 7.

19. Genetic sequence according to claim 14,
being the nucleotide sequence of Fig. 8.

20. Genetic sequence according to claim 14,
being the amino acid sequence of Fig. 8.

15 21. Genetic sequence according to claim 14,
being the nucleotide sequence of Fig. 9.

22. Genetic sequence according to claim 14,
being the amino acid sequence of Fig. 9.

23. Genetic sequence according to claim 14,
20 being the nucleotide sequence of Fig. 10.

24. Genetic sequence according to claim 14,
being the amino acid sequence of Fig. 10.

25. Genetic sequence according to claim 14,
being the nucleotide sequence of Fig. 11.

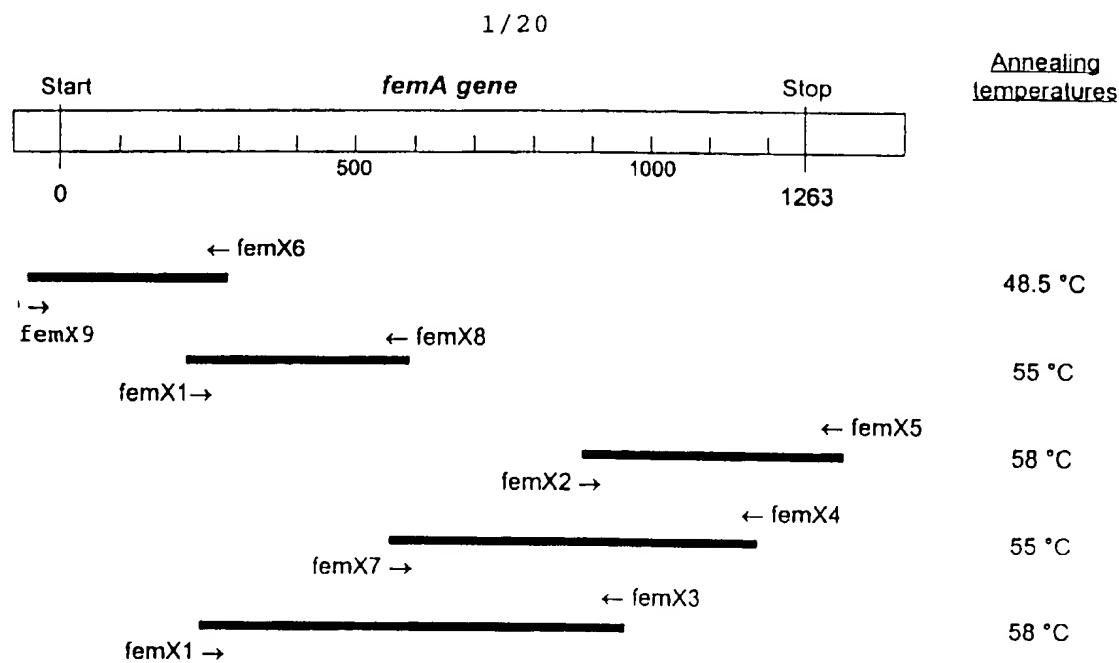
25 26. Genetic sequence according to claim 14,
being the amino acid sequence of Fig. 11.

27. Genetic sequence according to claim 14,
being the nucleotide sequence of Fig. 12.

28. Genetic sequence according to claim 14,
30 being the amino acid sequence of Fig. 12.

29. Genetic sequence according to claim 14,
being the nucleotide sequence of Fig. 13.

30. Genetic sequence according to claim 14,
being the amino acid sequence of Fig. 13.



Oligonucleotides

femX1	TTCMAATCGCGGTCCAGT	213-230
femX2	CAAGAACATGGCAACGAATTACC	913-935
femX3	TGGGTAAATTCGTTGCCATGTCT	937-915
femX4	CCAAGCATCTTCAGCATCTTC	1133-1113
femX5	TTCTTTAACTGTTAACTCTGTAATTTCA	1309-1281
femX6	ACATATTACTTAACTCGTTAAAGAA	290-265
femX7	CAGAAAAATGGTGTAAAGTAAGATT	559-585
femX8	AAGAAATCTTACTT TCACACCATTTTT	588-562
femX9	AACTCGAAAATAGAACTA	(-43)(-26)

FIG. 1

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	S. haemolyticus		S. hominis		S. aureus		S. epidermidis		S. saprophyticus		CONSENSUS	
a-c-t-ga	-	-	aa-taaaca	-	aa-tt-ct	-	aa-tt-tt	-	aa-tt-cc	-	ca-gt-g	-
a-t-t-gt	-	-	tg-gaaaact	-	aa-tt-tt	-	aa-tt-tt	-	aa-tt-tt	-	ca-gt-g	-
c-t-gt	-	-	tg-aggtat	-	aa-aa-cc	-	aa-aa-cc	-	aa-aa-cc	-	ca-gt-g	-
a-9t	-	-	tg-aggtac	-	gt-aa-tt	-	gt-aa-tt	-	gt-aa-tt	-	ca-gt-g	-
a-t-t-cat	-	-	aa-aa-cc	-	aa-aa-cc	-	aa-aa-cc	-	aa-aa-cc	-	ca-gt-g	-
a-t-t-at	-	-	ag-aagaca	-	aa-aa-cc	-	aa-aa-cc	-	aa-aa-cc	-	ca-gt-g	-
g-a-a-	-	-	tg-tt-tt	-	tt-aa-aa	-	tt-aa-aa	-	tt-aa-aa	-	ca-gt-g	-
-A-TA-GA-	-	-	TRGT-GG-AT	-	-AA-AA-AA-	-	-AA-AA-AA-	-	-AA-AA-AA-	-	-TG-T-T-T	-
T-TGTC	--A-----	--	GA-C-CA-	--	ATATA-GA-G	-	T-ATTGGC-GC	-	ATATA-GA-G	-	AC-GC-GT-C200	-

haemolyticus	ct- atcg -	-a-ac-t-	-a-c-cc-	t-t-	-a-----aa	-a-a-g--	c-aa--ccaa	-a-a-t-	-tagt--c-	t-t-tc-c
hominis	tc-tacta--	-a-att-a-	-t-t-ca	a-----t	-a-----at	-a-aa-gat-	-a-aa-ttc	-a-a-g-	-tagt--t	c-t-tc-a
aureus	tt-atctcg-	-a-ac-a-	-a-t-ta	a-----t	-a-----at	-a-ct-----gt	-a-ct-tgtc	-c-t-t-	-casa--t	c-c-tc-c
epidermidis	tt-atcg-	-a-ac-a-	-a-t-ta	g-----t	-g-----ct	-t-aa-t-----	-t-aa-t-----	-a-a-d-	-tagt--t	t-c-ta-a
saprophyticus	tt-aggtg-	-t-gc-g-	-a-aa-cc	c-----c	-a-----tt	-t-aa-a-----	-g-at-----tgca	-a-a-t-	-cgtt--t	t-t-ta-a
CONSENSUS	-T-----AA	GA-GA--T-C	C-AT-TT--G	-TCATT-ATG	GA-GATAC--	C-GA--C-AA	G-TT--	GAT-G-GA G A-----TT-TA	-TA-NA-G-700	

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FIG. 2b

NNNNNNNN NNNNNATGA ANTTTACNAA TTTAACNGCN ANAGANTNN GNNNNNTAC NGANNNWATG NCNNANAGNC ATTINACNA NANNNNNGNN
NANTANGNN TNAANTTGC NNANNNNNN GANNCCANN TAGTNGGGNAT NAANANRAN NATAANGANG TNATTGNGC NTGNNTNNN ACNGCNGTNC
CNGTNATGAA ANTNTNAAN TANTTTATT CNAANGNGG NCCNGTNATN GATTNTNANA ANNAGANTC NGTNCANTNN TTCTTTAANG ANTTNNNNAA
NTATNTNAA NANNNNNTN NNNTATANNT NNNNNNTNGAN CCNTANNTNN CNTATCAATA NNNAATCAT GANGGNANN TNNNNNGNAA TGNGCNGNN
GATTGGNTT TNGATNANNT NNNNNNNNTN GGNTNTYANC ANMINGGNTT NNNNANNGGN TTGANCNN TNNNNCAAT NNGNTNNCAN TCNGTNNTAN
NTTTANNNN NAAAANNNC NANGANNTNN TNAANNNAT GGATNGNNNT NGNAANNGNA ANACNAAAA AGTNNANAA ARTGGNGTNA AGTNMNTT
NNTNNNNAA CANGANNTNC CNATNTTNNG NTCAATNATG GANGATACNN CNGANICNA NGNNNTNNN GATNGNGANG ANNNNTNTA NTANANNGN
TNNNNNNATT NNAAGANN NGTNNTNGTN CCNTNTGCT ATATNNANT TGATGANTAN NTNNNNGAAN TNNNANNINGA NNGNNANN NTNNTAAAG
NNNNNNAA AGCANTNAAN GANATNGANA AANGCCNGA NANAANAN GCNNNAANA ANNNNNNAAN NNNTNNPANAN CAAANTNNING CNAANNNCA
AAAANNTNNN GANGNNNNN NNNTNNNAAN NNANCATGGN AANGAATTAC CNATNTCNGC NGNNNTNCCTN NTNATNAATC CNTNTGAAGT NGTNTPANTAN
GCNGGTGGNA CNTCNAATNN NTNNNGNCAN TTNGNGGNA GNTATGCNT NCANTGGNN ATGATTAANT ATGCNNTNNA NCATNNNATN NANNNGNTANA
ATTINTATGG NNTTAGNGGT NANTTTANNG ANGANGCNGA AGATGNNNGN GTNNNTNAANT TNAAAAANGG NTNNNNATGCN GANNTNNNTNG ANTANCTGG
NGANTTNNTN AAACCNATNA ANAANCNNN NTANNNNNN TATANNNCAN TNAAAANNT NNANNNNNANN NANNNNNNNA NANNNNNNNA NNNNNNNNN
NNNNNNATGA ATTACAG AGTTAANN

FIG. 3 CONSENSUS SEQUENCE

220 bases	<i>S.aureus</i>	<i>S.epidermidis</i>	<i>S. hominis</i>
<i>S.aureus</i>	-	-	-
<i>S.epidermidis</i>	17.7	-	-
<i>S.hominis</i>	13.2	16.8	-
<i>S.saprophyticus</i>	17.3	18.6	16.8

Base % (non appariated) between the primers bioU1 and bioU3

FIG4a

FIG.4b

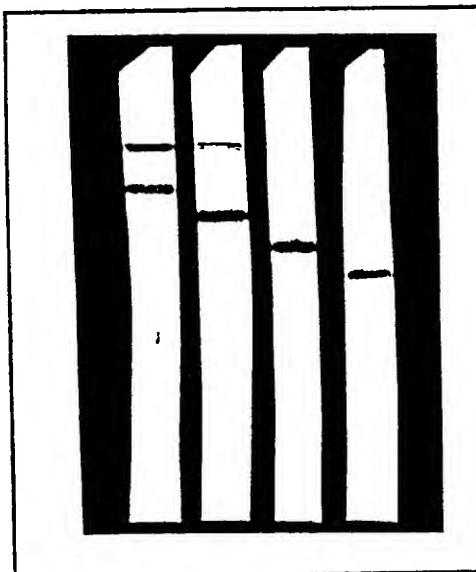
1 : *mecA*

2: *femA* *Sau*

3. *femA* *Sep*

4. *femA* *Sho*

5. *femA* *Ssa*



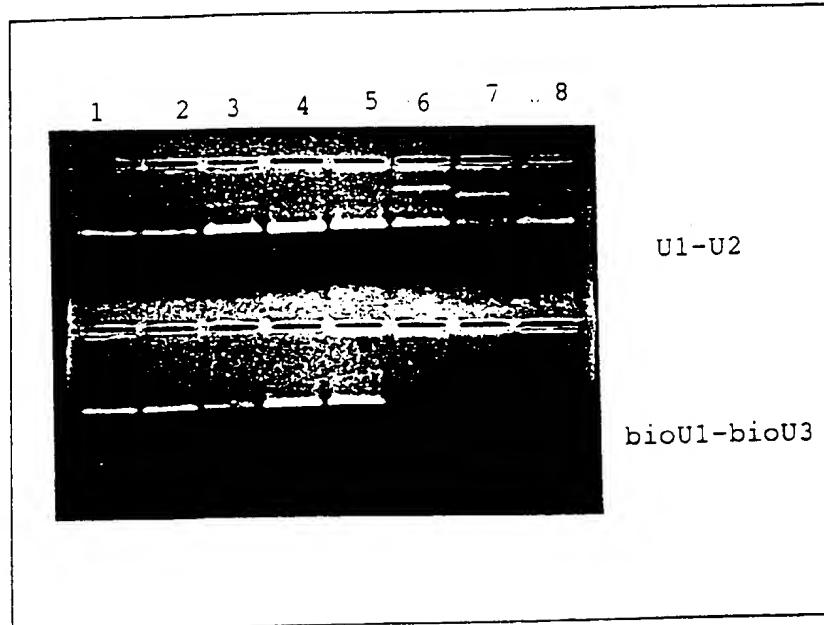


FIG. 5

AMPLIFICATION of CNS SPECIES under UNIVERSAL CONDITIONS.

(1) : *S. haemolyticus*
 (2) : *S. capitis*
 (3) : *S. cohnii* Th(reaction PCR) = 48°C
 (4) : *S. xylosus*
 (5) : *S. simulans*
 (6) : *S. lugdunensis*
 (7) : *S. schleiferi*
 (8) : *S. warneri*

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S. haemolyticus FIG. 6a

ATAATGAAGTTACAAATTACAGCTACAGAGTTGGCAATTACAGATAAGATGCCA
 MetLysPheThrAsnLeuThrAlaThrGluPheGlyAsnTyrThrAspLysMetPro
 70 90 110
 TATAGTCATTCACACAAATGACTGAAAATGAGATGAAAGTTGCAAATAAACAGAA
 TyrSerHisPheThrGlnMetThrGluAsnTyrGluMetLysValAlaAsnLysThrGlu
 130 150 170
 ACTCACTTAGTTGGTATAAAAATAAGATAATGAGGTTATTGCAGCCTGCATGTTGACA
 ThrHisLeuValGlyIleLysAsnLysAspAsnGluValIleAlaAlaCysMetLeuThr
 190 210 230
 GCAGTACCAAGTCATGAAATTTTTAACGAGCTTATTCTAACCGAGGACCTGTAATTGAT
 AlaValProValMetLysPhePheLysTyrPheTyrSerAsnArgGlyProValIleAsp
 250 270 290
 TATGATAATAGAGAGCTTGTTCACTTTTCTTAATGAGTTAACAAAGTATTAAACAG
 TyrAspAsnArgGluLeuValHisPhePheAsnGluLeuThrLysTyrLeuLysGln
 310 330 350
 CATAATTGTCTATATGTCGAGTTGACCTTATTACCATATCAATATTAAATCATGAT
 HisAsnCysLeuTyrValArgValAspProTyrLeuProTyrGlnTyrLeuAsnHisAsp
 370 390 410
 GGTGAAATTACAGGTAAATGCTGTAATGATTGGTTCTTGATAAGATGAAGCATCTCGGA
 GlyGluIleThrGlyAsnAlaGlyAsnAspTrpPhePheAspLysMetLysHisLeuGly
 430 450 470
 TTTGAACATGAAGGCTTACTAAAGGTTTGATCCGATTAAACAAATCCGATATCATTCT
 PheGluHisGluGlyPheThrLysGlyPheAspProIleLysGlnIleArgTyrHisSer
 490 510 530
 GTTTTAGATTAAAAAAATAAAACATCTAAAGATATATTAAATGGAATGGATAGTCTACGT
 ValLeuAspLeuLysAsnLysThrSerLysAspIleLeuAsnGlyMetAspSerLeuArg
 550 570 590
 AACGTAATACTAAAAAAAGTTCAAAAAAATGGTGTGAAAGTTAACGAAAGAAATTCCAAGAT
 LysArgAsnThrLysLysValGlnLysAsnGlyValLysValLysPheLeuSerGluGlu
 610 630 650
 GAACTCCAATCTCCGTTCATTTATGGAAGATAACCGAAACGAAAGAAATTCCAAGAT
 GluLeuProIlePheArgSerPheMetGluAspThrThrGluThrLysGluPheGlnAsp
 670 690 710
 AGAGATGATAGTTCTATTATAATCGCTATAGACATTCAAAGATCACGTGTTGACCA
 ArgAspAspSerPheTyrTyrAsnArgTyrArgHisPheLysAspHisValLeuValPro

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730 750 770
CTAGCTTATATTAAAGTTGATGAGTACATCGAAGAATTACAAATGAACGTGAAACTTTA
LeuAlaTyrIleLysPheAspGluTyrIleGluGluLeuGlnAsnGluArgGluThrLeu
790 810 830
AATAAAAGATGTTAATAAGCTTAAAAGATATTGAAAAACGACCAGACAATAAAAGGCA
AsnLysAspValAsnLysAlaLeuLysAspIleGluLysArgProAspAsnLysLysAla
850 870 890
TTTAATAAAAAGAAAATCTGAAAACAATTAGATGCCAATCAACAAAATTAGACGAG
PheAsnLysLysGluAsnLeuGluLysGlnLeuAspAlaAsnGlnGlnLysLeuAspGlu
910 930 950
GCTAAAAAATTACAAGCCGAAACATGGTAATGAATTACCAATTTCAGCAGGTTCTTCTT
AlaLysLysLeuGlnAlaGluHisGlyAsnGluLeuProIleSerAlaGlyPhePhePhe
970 990 1010
ATTAATCCATTGAAAGTTGTTATTATGCAGGTGGAACTTCTAATAATATAGACATT
IleAsnProPheGluValValTyrTyrAlaGlyGlyThrSerAsnLysTyrArgHisPhe
1030 1050 1070
GCAGGCAGTTATGCTATTCAATGGACAATGATTAACATGCAATTGATCATGGTATTGAT
AlaGlySerTyrAlaIleGlnTrpThrMetIleAsnTyrAlaIleAspHisGlyIleAsp
1090 1110 1130
AGATACAATTCTATGGTATTAGCGGTAATTAGTAGTGAAGACGCTGAAGATGTTGGAGTC
ArgTyrAsnPheTyrGlyIleSerGlyAsnPheSerGluAspAlaGluAspValGlyVal
1150 1170 1190
ATTAAATTAAAAAGGTTCAATGCAGACGTAATTGAGTATGTTGGAGACTTGAGAA
IleLysPheLysLysGlyPheAsnAlaAspValIleGluTyrValGlyAspPheValLys
1210 1230 1250
CCTATTAACAAACCTTGATTCAGTGTATAAGACACTCAAAAGATTAAAAAGATT
ProIleAsnLysProLeuTyrSerValTyrLysThrLeuLysIleLysLysArgPhe
1270 1290
AATTAAAGAGGGGAATAGACGAATATGAAATTACAGAGTTAAC
AsnEndArgGlyGluEndThrAsnMetLysPheThrGluLeuAsn

FIG. 6b

S. lugdunensisFIG. 7a

10 30 50

ACAGCAAATGAATTGGTGATTCACAGATCAAATGCCATATAGTCATTTACTCAAATG
 ThrAlaAsnGluPheGlyAspPheThrAspGlnMetProTyrSerHisPheThrGlnMet

70 90 110

ACAGGTTAACTATAATTAAAAGTTGCCAAAAACAGAACACATTAGTTGGTGTAA
 ThrGlyAsnTyrAsnLeuLysValAlaGluLysThrGluThrHisLeuValGlyValLys

130 150 170

AATAATAATAACGAAGTAATTGCAGCATGTTATTGACAGCTGTACCAAGTCATGAAGTT
 AsnAsnAsnAsnGluValIleAlaAlaCysLeuLeuThrAlaValProValMetLysPhe

190 210 230

TTTAAATACTTTACAGCAATAGAGGCCAGTTAGATTATGCTAACCAAGAACTTGTA
 PheLysTyrPheTyrSerAsnArgGlyProValIleAspTyrAlaAsnGlnGluLeuVal

250 270 290

CATTTTTCTTAATGAGCTAACTAAATATTAAAAAGTATACTGTCTCTATGTCCGC
 HisPhePhePheAsnGluLeuThrLysTyrLeuLysLysTyrAsnCysLeuTyrValArg

310 330 350

ATAGATCCATACTTACCTTATCAATATAGAGACCATGACGGTAATATAACGGCAAATGCT
 IleAspProTyrLeuProTyrGlnTyrArgAspHisAspGlyAsnIleThrAlaAsnAla

370 390 410

GGCAATGATTGGTTTTCAATAAAATGGAACAACTCGGATACCATCATGATGGCTTACA
 GlyAsnAspTrpPhePheAsnLysMetGluGlnLeuGlyTyrHisAspGlyPheThr

430 450 470

ACAGGATTGATCCAATATTACAAATCAGATTCCATTCTATTCTTAATTAAAGGATAAG
 ThrGlyPheAspProIleLeuGlnIleArgPheHisSerIleLeuAsnLeuLysAspLys

490 510 530

ACAGCTAAAGATTTAAATAATGGATAGTTACGTAAAAGAAATACCAAAAAAGT
 ThrAlaLysAspValLeuAsnAsnMetAspSerLeuArgLysArgAsnThrLysSer

550 570 590

TCAAAAAATGGAGTCAAAGTAAAGTTCTACTGAAGAAGAACTACCTATCTTCGTTCA
 SerLysAsnGlyValLysValLysPheLeuThrGluGluLeuProIlePheArgSer

610 630 650

TTTATGGAGCAGACGTCAGAATCTAAAGAATTCTCTGATAGAGACGACCAATTATTAC
 PheMetGluGlnThrSerGluSerLysGluPheSerAspArgAspAspGlnPheTyrTyr

670 690 710

AATCGGTTAACGACTATAAGATAGGGTGCTTGTGCCCTAGCATATTAAATTTGAT
 AsnArgPheLysTyrTyrLysAspArgValLeuValProLeuAlaTyrLeuLysPheAsp

10/20

730

750

770

GAATATATAGAAGAACTAACGAATGAACGCCACAAACTTAGAAAAAGATTAGGCAGCAAGCA
 GluTyrIleGluGluLeuThrAsnGluArgGlnThrLeuGluLysAspLeuGlyLysAla

790

810

830

CTTAAAGACATTGAGAAACGACCAAGATAACAAAAAGCTTATAATAAACGAGACAACCTA
 LeuLysAspIleGluLysArgProAspAsnLysLysAlaTyrAsnLysArgAspAsnLeu

850

870

890

CAACAAACAACCGATGCCAATCAACAAAAGTTAAATGAGGCTAACAGTTACAAGCGGAA
 GlnGlnGlnLeuAspAlaAsnGlnGlnLysLeuAsnGluAlaAsnGlnLeuGlnAlaGlu

910

930

950

CACGGTAATGAGTTACCTATCTCTGCCGGTTCTTATTATTAAATCCGTTGAAGTTGTA
 HisGlyAsnGluLeuProIleSerAlaGlyPhe^{II}IleAsnProPheGluValVal

970

990

1010

TACTACGCTGGAGGTACCGCTAATAAAATATCGTCATTGAGGTAGTTACGCCGTTCAAG
 TyrTyrAlaGlyGlyThrAlaAsnLysTyrArgHisPheAlaGlySerTyrAlaValGln

1030

1050

1070

TGGACTATGATTAACATATGCTATCGAACACGGCATAGACAGATATAATTCTACGGCATT
 TrpThrMetIleAsnTyrAlaIleGluHisGlyIleAspArgTyrAsnPheTyrGlyIle

1090

1110

1130

AGTGGAAACTTCTCAGATGATGCTGAAGACGCCAGGTGTCATTGCTTTAAAAAGGTTAT
 SerGlyAsnPheSerAspAspAlaGluAspAlaGlyValIleArgPheLysLysGlyTyr

1150

1170

1190

GGTGCAGAAGTGATTGAATACGTTGGTGATTTGTAAAACCTATAAAATAAACCTATGTAT
 GlyAlaGluValIleGluTyrValGlyAspPheValLysProIleAsnLysProMetTyr

1210

1230

1250

AAACTTTATTCACTGTTAAACGAATTCAAATAAGCTATAGAGGGAGAATGGATTAATT
 LysLeuTyrSerValLeuLysArgIleGlnAsnLysLeuEndArgArgMetAspEndLeu

1270

TGAAATTTACAGAGTTAAC
 EndAsnLeuGlnSerLeu

FIG. 7b

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S. xylosus

FIG. 8a

10 30 50
 ACGAAAAGAGTTGGGTGCATTCAGATAAAATGCCAATAGCCATTCACGCAAATG
 ThrGlnLysSerLeuGlyAlaPheSerAspLysMetProAsnSerHisPheThrGlnMet
 70 90 110
 GTAGGGAAATTATGAATTGAAAATTGCAGAAAGTACTGAAACACATTTAGTAGGTATAAAA
 ValGlyAsnTyrGluLeuLysIleAlaGluSerThrGluThrHisLeuValGlyIleLys
 130 150 170
 AACAAATGATAATGAAGTCATTGCAGCTTGTATTAACTGCAGTACCGAGTAATGAAATTG
 AsnAsnAspAsnGluValIleAlaAlaCysLeuLeuThrAlaValProValMetLysPhe
 190 210 230
 TTTAAGTATTTTATACTAATAGAGGTCCGGTTATAGATTTGAAAATAAGAATTAGTG
 PheLysTyrPheTyrThrAsnArgGlyProValIleAspPheGluAsnLysGluLeuVal
 250 270 290
 CATTACTTTCAATGAACTATCTAAATATGTGAAAAAACATAATGCGCTTTATTAAGA
 HisTyrPhePheAsnGluLeuSerLysTyrValLysLysHisAsnAlaLeuTyrLeuArg
 310 330 350
 GTTGATCCTTATTTAGCATATCAATACCGTAATCATGATGGTGAGGTATTGGAAAATGCA
 ValAspProTyrLeuAlaTyrGlnTyrArgAsnHisAspGlyGluValLeuGluAsnAla
 370 390 410
 GGACATGATTGGATTTCGATAAAATGAAGCAGCTGGATATAAACACCAAGGATTTTA
 GlyHisAspTrpIlePheAspLysMetLysGlnLeuGlyTyrLysHisGlnGlyPheLeu
 430 450 470
 ACTGGTTCGATTCAATTATTCAAATTAGGTTCCACTCTGACTGGATTAGTAGGTAAA
 ThrGlyPheAspSerIleIleGlnIleArgPheHisSerValLeuAspLeuValGlyLys
 490 510 530
 ACTGCTAAAGATGTACTAAATGGATAGTTACGTAAACGTAATACTAAAAAGTA
 ThrAlaLysAspValLeuAsnGlyMetAspSerLeuArgLysArgAsnThrLysLysVal
 550 570 590
 CAAAAAAATGGCGTGAAGTAAGGTTCTTAAGGGAAAGATGAGTTGCCATTTCGTTCA
 GlnLysAsnGlyValLysValArgPheLeuArgGluAspGluLeuProIlePheArgSer
 610 630 650
 TTCATGGAAGATACATCTGAAACTAAAGACTTGTACGATAGAGACGATGGCTTTACTAC
 PheMetGluAspThrSerGluThrLysAspPheAspAspArgAspGlyPheTyrTyr
 670 690 710
 AATAGATTAAGGTATTATAAGATCGCGTATTAGTACCTCTAGCTTATATGGATTCAAT
 AsnArgLeuArgTyrTyrLysAspArgValLeuValProLeuAlaTyrMetAspPheAsn

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730 750 770
 GAATATATTGAAGAATTGCAAGCTGAACTGAGGTGTTAAGCAAAGATATCAATAAGCA
 GluTyrIleGluGluLeuGlnAlaGluArgGluValLeuSerLysAspIleAsnLysAla
 790 810 830
 GTAAAAGATATCGAGAAAAGACCTGAAAATAAAAAGCATATAATAAAAAAGATAATCTA
 ValLysAspIleGluLysArgProGluAsnLysLysAlaTyrAsnLysLysAspAsnLeu
 850 870 890
 GAGAAACAACCTTATAGCGAACATCAACAAAAATTGATGAAGCTAAACTCTACAAGAGAAG
 GluLysGlnLeuIleAlaAsnGlnGlnLysIleAspGluAlaLysThrLeuGlnGluLys
 910 930 950
 CATGGTAACGAACCTACCAATCTCAGCAGCATATTCATCATTAACCTTATGAAGTAGTG
 HisGlyAsnGluLeuProIleSerAlaAlaTyrPheIleIleAsnProTyrGluValVal
 970 990 1010
 TATTATGCGGGTGGAACGTCAAATGAGTTAGACATTTGCTGGTAGTTATGCCATTCAA
 TyrTyrAlaGlyGlyThrSerAsnGluPheArgHisPheAlaGlySerTyrAlaIleGln
 1030 1050 1070
 TGGAAGATGATTAACATGCTATTGACCATAATATTGATAGATATAATTTTATGGAATT
 TrpLysMetIleAsnTyrAlaIleAspHisAsnIleAspArgTyrAsnPheTyrGlyIle
 1090 1110 1130
 AGTGGTCATTTACAGAAGATGCAGAAGATGCCGGTAGTTAAATTAAAAAGGATT
 SerGlyHisPheThrGluAspAlaGluAspAlaGlyValValLysPheLysLysGlyPhe
 1150 1170 1190
 AATGCGGATGTAGTGGAAATATGTTGGTAGTTATTAAACCAATCAATAACCAATGTAC
 AsnAlaAspValValGluTyrValGlyAspPheIleLysProIleAsnLysProMetTyr
 1210 1230 1250
 AAAATTATACGACATTAAAGAAAATTAAAGATAAAAGAAATAAACATTAAATAGAAGG
 LysIleTyrThrThrLeuLysLysIleLysAspLysLysLysEndThrPheAsnArgArg
 1270 1290
 GAACTAAGCTAGAATGAAATTACAGAGTTAAACC
 GluLeuSerEndAsnGluIleTyrArgValLys

FIG. 8b

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S. capititis FIG. 9a

10	30	50
ACAGCTAAAGAATTAGTAGCTTACTGATCAAATGCCCTATAGCCATTTACTCAGATG ThrAlaLysGluPheSerAspPheThrAspGlnMetProTyrSerHisPheThrGlnMet		
70	90	110
GAAGGTTAATTATGAACTTAAAGTTGCTGAAGGTACGGATTACATCTCGTAGGAATTAAA GluGlyAsnTyrGluLeuLysValAlaGluGlyThrAspSerHisLeuValGlyIleLys		
130	150	170
AATAATGACAACCAAGTGATTGCAGCATGTTTATTAACACTGCTGTACCTGTAATGAAAATT AsnAsnAspAsnGlnValIleAlaAlaCysLeuLeuThrAlaValProValMetLysIle		
190	210	230
TTTAAATATTTTACTCAAATCGCGGGCCAGTGATTGATTATGATAATAAGAGCTTGT PheLysTyrPheTyrSerAsnArgGlyProValIleAspTyrAspAsnLysGluLeuVal		
250	270	290
CACTTTCTTAATGAATTAAAGTAAATATGAAAAAGCATAATTGCTTTATCTAAGA HisPhePhePheAsnGluLeuSerLysTyrValLysLysHisAsnCysLeuTyrLeuArg		
310	330	350
GTTGACCCTTATCTTCCTTATCAATACTAAATCATGACGGTGAATTATTGGAAATGCT ValAspProTyrLeuProTyrGlnTyrLeuAsnHisAspGlyGluIleIleGlyAsnAla		
370	390	410
GGCCATGATTGGTTTTCAATAAGATGGAAGAATTAGGATTGAAACATGAAGGCTTCAT GlyHisAspTrpPhePheAsnLysMetGluGluLeuGlyPheGluHisGluGlyPheHis		
430	450	470
AAAGGCTTCCATCCTATCTTACAAGTAAGATATCATTGAGTTAGATTAAAGATAAAA LysGlyPheHisProIleLeuGlnValArgTyrHisSerValLeuAspLeuLysAspLys		
490	510	530
ACGGCTAAAGATGACTCAAAGGAATGGATAGTTAAGAAAGCGTAATACTAAGAAAGTA ThrAlaLysAspValLeuLysGlyMetAspSerLeuArgLysArgAsnThrLysLysVal		
550	570	590
CAAAAAAAATGGTGTCAAAGTCGTTCCATCCGAAGATGAATTACCTATCTTAGATCA GlnLysAsnGlyValLysValArgPheLeuSerGluAspGluLeuProIlePheArgSer		
610	630	650
TTTATGGAAGATACTACAGAAACGAAAGAGTTCGCCGATAGAGATGATAGTTCTATTAT PheMetGluAspThrThrGluThrLysGluPheAlaAspArgAspAspSerPheTyrTyr		

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670	690	710
AATCGATTAAAATACTTTAAAGATAGAGTATTAGTACCATAGCATATGTTGACTTCGAT		
AsnArgLeuLysTyrPheLysAspArgValLeuValProLeuAlaTyrValAspPheAsp		
730	750	770
GAGTATATTGAAGAACCTAATAATGAAAGAGATGTTCTTAATAAGATTAAATAAGGCG		
GluTyrIleGluGluLeuAsnAsnGluArgAspValLeuAsnLysAspLeuAsnLysAla		
790	810	830
CTCAAAGATATTGAGAACGACTGATAATAAGAAAGCTTATAACAAAAGAGATAATCTT		
LeuLysAspIleGluLysArgProAspAsnLysAlaTyrAsnLysArgAspAsnLeu		
850	870	890
CAACAACAATTAGATGCAAATCAACAAAAATTGATGAAGCTAAAAACTTACAACAAGAA		
GlnGlnGlnLeuAspAlaAsnGlnGlnLysIleAspGluAlaLysAsnLeuGlnGlnGlu		
910	930	950
CATGGTAATGAATTACCTATTTCAGCTGGATATTCTTCATTAATCCGTTGAAGTTGTT		
HisGlyAsnGluLeuProIleSerAlaGlyTyrPhePheIleAsnProPheGluValVal		
970	990	1010
TATTACGCAGGTGGCACATCGAATCGTTATCGTCACTATGCCGGAAGTTATGCAATTCAA		
TyrTyrAlaGlyGlyThrSerAsnArgTyrArgHisTyrAlaGlySerTyrAlaIleGln		
1030	1050	1070
TGGAAAATGATAAACTATGCTTAGAACATGGAATTACCGTTATAATTGAGTT		
TrpLysMetIleAsnTyrAlaLeuGluHisGlyIleAsnArgTyrAsnPheTyrGlyVal		
1090	1110	1130
AGTGGGGACTTCAGTGAAGACGCTGAAGATGTAGGAGTAATTAGTTCAAGCTATGCTAA		
SerGlyAspPheSerGluAspAlaGluAspValGlyValIleLysPheLysLysGlyTyr		
1150	1170	1190
AATGCTGATGTTATTGAATATGTAGGTGATTTATCAAGCCAATCAATAAACCTATGTAT		
AsnAlaAspValIleGluTyrValGlyAspPheIleLysProIleAsnLysProMetTyr		
1210	1230	1250
GCAATCTATAACGCACTTAAAAAGTTAAAGAAATAGATTTTACCAACCCAATTATCT		
AlaIleTyrAsnAlaLeuLysLysLeuLysLysEndIlePheLeuProThrGlnLeuSer		
1270		
AATTATGAAATTACAGAGTTAA		
AsnTyrGluIleTyrArgVal		

FIG. 9b

10

30

FIG.10a
50ACGACGGCTGAATTGGTGCCTTACAGATCAAATGCCATATAGCCATTCACGCAAATG
ThrThrAlaGluPheGlyAlaPheThrAspGlnMetProTyrSerHisPheThrGlnMet

70

90

110

GTAGGGAACATATGAATTAAAGGTTGCTGAAGGTGTTGAAACACATCTTGTGCGCATTAAA
ValGlyAsnTyrGluLeuLysValAlaGluGlyValGluThrHisLeuValGlyIleLys

130

150

170

GATAACAACAATAACGTACTAGCAGCATGTTACTGACAGCAGTGCCAGTAATGAAGTTT
AspAsnAsnAsnAsnValLeuAlaAlaCysLeuLeuThrAlaValProValMetLysPhe

190

210

230

TTTAAATATTTTATTCAAACCGCGGACCACTGACTACGAAAATAAAGAGCTCGTT
PheLysTyrPheTyrSerAsnArgGlyProValMetAspTyrGluAsnLysGluLeuVal

250

270

290

CATTCTTTTAATGAACCTTCAAAATATGTTAAGAAATATCACGCATTGTATTCAGA
HisPhePheAsnGluLeuSerLysTyrValLysLysTyrHisAlaLeuTyrLeuArg

310

330

350

GTAGACCCATTACCAATGTTAAAGCGAAACCATGATGGTGAAGTGATTGAAAGATAAC
ValAspProTyrLeuProMetLeuLysArgAsnHisAspGlyGluValIleGluArgTyr

370

390

410

GGCAGTGACTIONTTTGATAAAATGGCTGAATTAAACTTGAACATGAAGGTTTCACA
GlySerAspTrpPhePheAspLysMetAlaGluLeuAsnPheGluHisGluGlyPheThr

430

450

470

ACTGGGTTTGATACAATAAGGCAAATTCGTTTCATTCTGTGCTCGATGTTGAAAATAAA
ThrGlyPheAspThrIleArgGlnIleArgPheHisSerValLeuAspValGluAsnLys

490

510

530

ACATCAAAAGACATCTAAATCAAATGGATAATTAAAGGAAAAGAAAATACGAAAAAAAGTA
ThrSerLysAspIleLeuAsnGlnMetAspAsnLeuArgLysArgAsnThrLysLysVal

550

570

590

CAGAAAAATGGGTGAAAGTCCGCTATCTAAACGAAGATGAATTACATATTTCCGTTCG
GlnLysAsnGlyValLysValArgTyrLeuAsnGluAspGluLeuHisIlePheArgSer

610

630

650

TTTATGGAAGATACTCTGAAACAAAAGATTGAGATAGAGATGACGATTTTATTAT
PheMetGluAspThrSerGluThrLysAspPheValAspArgAspAspPheTyrTyr

670

690

710

CATCGTATGAAATACTATAAGATCGTGTCCCGTACCACTAGCGTATATTGATTTAAAT
HisArgMetLysTyrTyrLysAspArgValArgValProLeuAlaTyrIleAspPheAsn

16/20

730

750

770

GCATATTTAGCAGAGCTAACACTGAAGCGCAAGACTTTAAAAAGAAATTGCAAAAGCA
 AlaTyrLeuAlaGluLeuAsnThrGluAlaGlnAspPheLysLysGluIleAlaLysAla

790

810

830

GATAAAGACATCGACAAGCGTCTGAAAATCAGAAAGCCATAAATAAAAAGAAAAATTAA
 AspLysAspIleAspLysArgProGluAsnGlnLysAlaIleAsnLysLysLysAsnLeu

850

870

890

GAGCAACAACTAGAACGAACTAACAGCTAAAATAAAAGAACAGAAACATTGCAACTTAA
 GluGlnGlnLeuGluAlaAsnGlnAlaLysIleLysGluAlaGluThrLeuGlnLeuLys

910

930

950

CACGGTGACACATTACCGATTCGGCTGGATTCTTATTATTAAATCCATTGAGGTTGTT
 HisGlyAspThrLeuProIleSerAlaGlyPhePheIleIleAsnProPheGluValVal

970

990

1010

TATTATGCAGGCGGCACAGCAAACGAATTCGTCATTTGCTGGAGCTACGCAGTGCAA
 TyrTyrAlaGlyGlyThrAlaAsnGluPheArgHisPheAlaGlySerTyrAlaValGln

1030

1050

1070

TGGGAAATGATTAATTATGCGATTGATTATCAAATTCAAAGATATAACTTTATGGCATT
 TrpGluMetIleAsnTyrAlaIleAspTyrGlnIleProArgTyrAsnPheTyrGlyIle

1090

1110

1130

AGTGGTGATTTCAGAACGATGCAAGAGATGCAGGTGTGATAAAATTAAAAAGGCTAT
 SerGlyAspPheSerGluAspAlaGluAspAlaGlyValIleLysPheLysLysGlyTyr

1150

1170

1190

AATGCAGAACGTAATAGAATATGCGGTGATTTATTAAGCCTATAAACAAACCTGCCTAT
 AsnAlaGluValIleGluTyrValGlyAspPheIleLysProIleAsnLysProAlaTyr

1210

1230

1250

ACAGTCTACTAAAATTAAAGCAATTAAAGACAAGATAAAAGATAAGATATAGCAAAG
 ThrValTyrLeuLysLeuLysGlnLeuLysAspLysIleLysArgEndAspIleAlaLys

1270

1290

AGAAGGGGATTATTGGTATGAAATTACAGAGTTAA
 ArgArgGlyPheIleGlyMetLysPheThrGluLeu

FIG.10b

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S. sciuriFIG.11a

10 30 50

ACACTGGAATTTGAAGCTTTACAAATAAAATGCCGTACGCGCATTTACACAAGCAGTA
ThrLeuGluPheGluAlaPheThrAsnLysMetProTyrAlaHisPheThrGlnAlaVal

70 90 110

GGTAATTATGAATTAAAAACATCTGAAGGTACTTCAACACATTTAGTAGGGGTCAAAGAT
GlyAsnTyrGluLeuLysThrSerGluGlyThrSerThrHisLeuValGlyValLysAsp

130 150 170

AATCAAGGTGAAGTATTAGCTGCGTGTCTGTTAACAAAGTGTACCAGTTATGAAGAAATT
AsnGlnGlyGluValLeuAlaAlaCysLeuLeuThrSerValProValMetLysLysPhe

190 210 230

AATTACTTTACTCAAATAGAGGACCAGTAATGGATTATGACAACAAAGAACTTGTTGAC
AsnTyrPheTyrSerAsnArgGlyProValMetAspTyrAspAsnLysGluLeuValAsp

250 270 290

TTTTCTTAAAGAAATCGTGAGCTATTTAAAAAGTTATAAGGATTATTCTTCTAGAAC
PhePhePheLysGluIleValSerTyrLeuLysSerTyrLysGlyLeuPhePheArgIle

310 330 350

GATCCTTACTTGCCATATCAACTAACAGAGATCATGATGGCAATTAAAAATCATTCAAC
AspProTyrLeuProTyrGlnLeuArgAspHisAspGlyAsnIleLysLysSerPheAsn

370 390 410

CGTGATGGTTAATTAAACAATTGAATCATTAGGTTATGAACACCAAGGCTTCACA
ArgAspGlyLeuIleLysGlnPheGluSerLeuGlyTyrGluHisGlnGlyPheThrThr

430 450 470

GGTTTCCACCCAATACATCAAATTAGATGGCATTCTGTACTTGATTAGAAAGTATGGAC
GlyPheHisProIleHisGlnIleArgTrpHisSerValLeuAspLeuGluSerMetAsp

490 510 530

GAAAAAGACGCTCATCAAGAACATGGACAGTTAACAAAAAGAAATACTAAAAAGTTCAA
GluLysThrLeuIleLysAsnMetAspSerLeuArgLysArgAsnThrLysLysValGln

550 570 590

AAAAATGGTGTAAAGTTCGTTCTATCTAAAGATGAAATGCCGATATTCCGTCAATT
LysAsnGlyValLysValArgPheLeuSerLysAspGluMetProIlePheArgGlnPhe

610 630 650

ATGGAAGATACTACAGAGAAGAAAGATTCAACGATCGTGGCGATGACTTCTATTACAAT
MetGluAspThrThrGluLysLysAspPheAsnAspArgGlyAspAspPheTyrTyrAsn

18/20

670

690

710

AGATTAAAATACTTGTAAAATGTAAAGATTCTTCTTACATATAGACTTGTAAACTTAC
 ArgLeuLysTyrPheGluAsnValLysIleProLeuAlaTyrIleAspPheGluThrTyr

730

750

770

ATTCCACAATTAGAAAAAGAACATGAACAAATACAACAAAGATATTGCAAAAGCTGAAAAA
 IleProGlnLeuGluLysGluHisGlnTyrAsnLysAspIleAlaLysAlaGluLys

790

810

830

GATTTAGAAAAGAAACAGATAATCAAAAACGATTATAAAATAGACAACCTAAACAA
 AspLeuGluLysLysProAspAsnGlnLysThrIleAsnLysIleAspAsnLeuLysGln

850

870

890

CAAAGAGAAGCAAATGAAGCTAAATTAGAAGAAGCACTTCAACTACAACAAGAACATGGT
 GlnArgGluAlaAsnGluAlaLysLeuGluAlaLeuGlnLeuGlnGluHisGly
 !!

910

930

950

GATACATTACCAATAGCAGCTGGTTCTTATTATAATCCATTGAAGTTGTATATTAT
 AspThrLeuProIleAlaAlaGlyPhePheIleIleAsnProPheGluValValTyrTyr

970

990

1010

GCAGGTGGTTCATCGAATGAATATCGTCACTTGCAGGTAGTTATGCAATTCACTGGAA
 AlaGlyGlySerSerAsnGluTyrArgHisPheAlaGlySerTyrAlaIleGlnTrpGlu

1030

1050

1070

ATGATTAAATACGGCTTAGATCACAAACATTGACCGTTATAACCTATGGTATCAGCGGA
 MetIleLysTyrAlaLeuAspHisAsnIleAspArgTyrAsnPheTyrGlyIleSerGly

1090

1110

1130

GACTTCTCAGAAGATGCACCTGATGTTGGCGTTATTAAATTAAAAAGGTTACAATGCA
 AspPheSerGluAspAlaProAspValGlyValIleLysPheLysLysGlyTyrAsnAla

1150

1170

1190

GATGTTTATGAATATATTGGTGTTCGTTAACCAATTAAATAAAACCAGCGTACAAAGCA
 AspValTyrGluTyrIleGlyAspPheValLysProIleAsnLysProAlaTyrLysAla

1210

1230

1250

TATACAAACACTAAAAAGTATTAAAAATAAAATGATTTCAAGTAAGAGAGGAATTAG
 TyrThrThrLeuLysLysValLeuLysLysEndMetIlePheSerLysArgGlyIleEnd

1270

ATAATATGAAATTACAGAGTTAA
 IleIleEndAsnLeuGlnSerEnd

FIG.11b

Staphylococcus hominis

taaaaaattttaaatttagtcaactcaaattaaataaaggatttcttaaaatttaggatttatagagataATGAAAGTTTACAAATTAAACAGCTACAGAAATTGGCG 100
 F T E K M P Y S H F T Q M T E N Y E L K V A E K T E T H L V G I K
 ATTACTGAAAAATGCCATATAGCCATTACAGATGACTGAAATTATGAGTTAAAGTGCTGAGAAAACACTGAAACTCATTTAGTAGGAAATTAA 200
 N K D N E V I A A C M L T A V P V M K I P K Y F Y S N R G P V I D
 AAATAAAGATAATGAAAGTCATGCTGCTTCTATGCTAACTGCTGACCGTTATGAAAATTTTAAATTTTAAATTTTAAATCAATTCTGGTCAGTCATTGAT 300
 Y E N K E L V H F F N E L S K Y L K Q Q H C L Y V R I D P Y L P Y
 TATGAAAACAAAGAAACTCGTTCACTTTTAAACAAACATTATGTAAGTAAATTAAGTAAATTAACGAAATTTCATGTTATATGTAAGTATAGACCCCTTATTGGCCTT 400
 M K F T N L T A T E F G D
 ATCAATATCGTAAATCATGATGGTGTGATATTACAGGAATGGAAATGGTTCTCGATAAATGAAAACATTAGGATAATGGATAATGAAAGATACTACAG 500
 Q Y R N H D G D I T G N A G N D W F F D K M K Q L G Y Q H E G F T
 ATCAATATCGTAAATCATGATGGTGTGATATTACAGGAATGGAAATGGTTCTCGATAAATGAAAACATTAGGATAATGGATAATGAAAGATACTACAG 600
 AACAGGATTTCATCCAATATTACAAATTACAAATTGGTTCATTCAGTTAAATTAAAGGATAAAACTGCTAAAGATGTATAATGGATAATGGATAATGTTAACGA
 T G F D P I L Q I R F H S V L N L K D K T A K D V L N G M D S L R
 AAAAGGAAATACTCAAAAAAAATGGTGTAAAGTAAGATTTCATCAAAAGAAATTACCTTACATGATCATTTCAGATCATTATGGAAAGATACTACAG 700
 K R N T K V Q K N G V K V R F L T K E E L P I F R S F M E D T S E
 AGACTAAAGAATTTCCTGATAGAGGATAGTGTGATCATTTAAAGCATAGTGTATAATCGATTGATCATTTAAAGCATAGTGTATAATGGATAATGGATAATGTA 800
 T K E F S D R E D S F Y Y N R F D H F K D R V L V P L A Y I K F D
 TGAATATCTTGAAGAACTTCATGCGAACCGTCAGACATTAATTAAGGACTTAAACAAAGCTCTAAAGGATATTGAAAAACGACAGATAACAAAAAGCA 900
 E Y L E E L H A E R Q T L N K D L N K A L K D I E K R P D N K K A
 CAAATAAAAAATAAATTAGAACAGCAATTAAAGCAAAATGAGCAAAAAATTGATGAAGCAACACAACTTCATAATTAGAACATGGTACGAAATTACAA 1000
 Q N K K I N L B Q Q L K A N E Q K I D E A T Q L Q L E H G N E L P I
 TATCTGCTGGATTCCTCTTAAATTATGCCATTGAACTGTTATATTATGCCAGGTCAAATAAATATGACACACTCGCTGGAAAGTTATGCGAGTC 1100
 S A G F F P I N P F E V V Y A G G T S N K Y R H F A G S Y A V Q
 ATGGACTATGATAATTATGCCATTGATCATGGCATTGACCGTTATAATTGAGTTATGGATTAGTGGTCATTTCAGATGATGCTGAGATGGAGGTGTT 1200
 W T M I N Y A I D H G I D R Y N F Y G I S G H F T D D A E D A G V
 GTAAAATTTAAAGGATTAAATGCAAGATGTAATTGAAATGTTGGTAAACCTATAAAACCAATGTTCACTATAACACCTTA 1300
 V K F K K G F N A D V I E Y V G D F V K P I N K P M Y S L Y T T L K
 AAAATAAATTAAAGGAGATTGAAATTAAAGggggaaatagtggaaa 1343
 K I K K R L N // /

FIG. 12

Staphylococcus saprophyticus

FIG. 13.

1371

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European Patent
Office

EUROPEAN SEARCH REPORT

Application Number
EP 00 87 0127

DOCUMENTS CONSIDERED TO BE RELEVANT			CLASSIFICATION OF THE APPLICATION (Int.Cl.7)
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	
X	WO 96 07917 A (NANOGEN) 14 March 1996 (1996-03-14)	17-20	C12Q1/68
Y	* the whole document * ---	1-16	
Y	ARMOUR ET AL: "MEASUREMENT OF LOCUS COPY NUMBER BY HYBRIDISATION WITH AMPLIFIABLE PROBES" NUCLEIC ACIDS RESEARCH, GB, OXFORD UNIVERSITY PRESS, SURREY, vol. 28, no. 2, 2000, pages 605-609, XP002138423 ISSN: 0305-1048 * the whole document * ---	1-16	
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A	EP 0 420 260 A (HOFFMANN LA ROCHE) 3 April 1991 (1991-04-03) * the whole document * ---	---	
The present search report has been drawn up for all claims			
Place of search	Date of completion of the search	Examiner	
THE HAGUE	12 December 2000	Hagenmaier, S	
CATEGORY OF CITED DOCUMENTS		T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons & : member of the same patent family, corresponding document	
X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document			